The Bryophyte Flora of the Big Thicket National Preserve: 
Hardin, Tyler, and Polk Counties, Texas

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ABSTRACT

The Big Thicket region is a complex mix of terrestrial and aquatic ecosystems which produce a very diverse flora and fauna. A floristic inventory of the bryophyte flora in the Big Thicket National Preserve, Hardin, Polk, and Tyler Counties, Texas, was conducted from January 2007 to September 2011. This inventory resulted in an updated checklist of 179 species of hornworts, liverworts, and mosses, in 98 genera and 54 families. Thirteen potentially new state records, twelve (12) liverworts and one (1) moss, are reported.

INTRODUCTION

The “big thicket” region of southeast Texas has long been referred to as a biological crossroads. Situated as the intersection of several distinct eco-regions; the area harbors a unique mix of plants and animals indicative of the southeastern swamps, eastern forests, central plains, and the southwestern United States. Its biota includes species from tropical and subtropical biomes, as well as those characteristic of the arid west (Gunter 1993). The region also represents the western-most extension of the southeastern evergreen forests (McLeod 1971), and is the western boundary for distributions of many aquatic insects with largely eastern affinities (Abbott et al. 1997). The Big Thicket National Preserve (BITH), founded in 1974, seeks to preserve the fragmented remnants of a once much larger and contiguous region of natural ecosystems. Comprising nearly 105,000 acres in a disparate mix of land units, and several interconnecting water corridors, BITH was the first unit of the National Park Service (NPS) to be set aside with the expressed intent of protecting biodiversity.

Surveys of vascular plants have been reported for most regions of Texas, and in particular, east Texas. However, many of the earliest east Texas reports (Gow 1904, Parks and Cory 1938) made no mention, or only a passing reference to bryophytes, and none provide real inventories or checklists of these groups. Even reports from surveys conducted much later (McLeod 1971) neglect to include bryophytes as part of their inventories. Following the establishment of the BITH, a broad survey of plant and fungal groups within the newly established preserve was initiated (Schedlbauer 1974). This report included what could be considered the first inventory of bryophytes within the legal confines of the BITH. Mueller 1974, reported 77 species in 49 genera as occurring in selected units of the preserve. Following these initial reports, additional authors have supplemented the known bryophyte flora of the BITH (Stoneburner and Wyatt 1979, Lodwick and Snider 1980, Bazan 1980, Wyatt, et al. 1980, Reese 1984). Despite these subsequent reports, a number of BITH’s operational units have limited, or no, reports of bryophytes.

The objective of this inventory was to provide an overall inventory of extant bryophyte taxa within the entirety of the BITH. In the context of this inventory, bryophytes include taxa from the Bryophyta (moss), Marchantiophyta (liverwort), and Anthocerotophyta (hornwort) phyla.
METHODS

Field surveys were conducted in each of the nine (9) land units: Beaumont, Beech Creek, Big Sandy Creek, Canyonlands, Hickory Creek Savannah, Jack Gore Baygal, Lance Rosier, Lobolly, and Turkey Creek. Due to continuing access problems following Hurricane Rita in 2005, most the length of the corridor units (Upper and Lower Neches River, Big Sandy Creek, Menard Creek, Pine Island Bayou) was excluded from this survey. While in the field, surveys were conducted utilizing a modified floristic habitat sampling method. This method as described by Newmaster, et al. 2005, focuses on a stratification of different mesohabitats into their component microhabitats for sampling bryophytes and was shown to produce a significant increase (~50%) in the number of taxa documented over the standard plot sampling method. Marks and Harcombe 1981, defined the vegetation types within the Big Thicket region of east Texas based generally on topographic position and the dominant over-story vegetation. Each of these is further subdivided into a number of communities such as: sandhill pine forest, upland pine forest, wetland pine savannah, upper slope pine-oak forest, wetland baygall shrub thicket, swamp cypress tupelo forest. Within each of these communities, bryophytes inhabit a variety of substrates such as: soil and rock along stream banks, decaying or living tree bases, specialized soil types, specific tree species, etc.

Sampling locations were spatially distributed to include a minimum of one (1) location within each mesohabitat and microhabitat encountered in each of the individual land units. Within each microhabitat the various substrates were sampled for bryophytes. Sampling trips were conducted throughout the year, with an increased emphasis in the late fall and early spring to document taxa which are typically ephemeral in nature. Collections were transported to the S. M. Tracy Herbarium (TAES) for processing, identification, and vouchering. Identifications were made using various references, however, the primary sources utilized were Briel 1970, and Schuster 1966, 1969, 1974, 1980, 1992, 1992a for the Anthocerotophyta and Marchantiophyta; Crum and Anderson 1981, and Reese 1984 for the Bryophyta. Nomenclature follows ITIS 2011 for all taxa with the following exceptions: Scapania nemorosa fo. whitehouseae R.M. Schust., follows Schuster 1992a; Ctenidium molluscum (Hedw.) Mitt. and Riccia stenophylla Spruce, follow USDA 2011. Identifications were made by Dale A. Kruse and Paul V. Roling. A limited number of specimens were sent to Paul G. Davison for identification as indicated in the accompanying database files. Identifications from previously published reports were not independently verified. All data has been entered into a project data file which has been submitted to the NPS. Vouchers are deposited at TAES under an existing NPS repository agreement. Specimens are stored in archival quality bryophyte packets with an accompanying label containing all relevant collection information.

In addition to field collections, a survey of selected state and regional herbaria was conducted to determine where additional BITH collections may be housed. Appropriate specimen data was obtained from five (5) herbaria: ASTC, BRIT, SHST, TEX/LL, and MO (W.R. Reese’s collections transferred from LAF). Specimens from BAYLU were not made available for inspection (acronyms follow Thiers, continuously updated). Specimen identifications obtained from these herbaria were not verified. The data acquired from these herbaria is included in a second historical records data file.

RESULTS

Field work was conducted from January 2007 through September 2011 and resulted in the accumulation of approximately 693 voucher samples from the BITH. Historical data, obtained from searches of the herbaria listed in the previous section, added another 730 records for Hardin, Polk, and Tyler Counties. However, only 217 could be reliably attributed to the BITH, the remainder could not be verified as BITH collections and are provided as additional county records.

A cumulative checklist of bryophytes within the BITH was compiled from collections made in the course of this inventory, records from previously published reports of taxa not found in this inventory, and additional records obtained from the search of herbaria. The current checklist of bryophytes in the BITH
comprises 179 species, in 98 genera and 54 families. The total reported taxa consist of 3 hornworts, 69 liverworts, and 107 mosses. The families with the most species were Lejeuneaceae (16 spp.), Fissidentaceae (9 spp.), Hypnaceae (8 spp.), Polytrichaceae, Pottiaceae, Ricciaceae, Sphagnaceae (each with 7 spp.), and Funariaceae, Jubulaceae (each with 6 spp.). Species-rich genera included Fissidens (9 spp.), Sphagnum (7 spp.), Frullania, Riccia (each with 6 spp.), Cololejeunea (5 spp.), and Funaria, Lejeunea, Leskea, Leucolejeunea, Riccardia (each with 4 spp.).

The current checklist includes several possible new records for the state of Texas. Based on the current inventory, and previously published reports, thirteen (13) taxa are potentially new to the state. Four (4) of these (Cololejeunea biddlecomiae (Aust.) Evans, Cylindrocolea rizantha (Mont.) Schust., Jungermannia gracililima Sm., Metzgeria unigera A. Evans) were added to the checklist based on records from a Master’s Thesis (Bazan 1980). At this time no reports have been found of these taxa in the literature and vouchers for these specimens have not been located. Therefore, the listing of these taxa is considered tentative until vouchers can be located and identifications verified. The remaining eleven (11) taxa are anticipated new records for the state (Cephaloziella hyalina Douin, Ceratolejeunea laetefusca (Aust.) Schust., Cololejeunea contractiloba Evans, Cololejeunea setiloba Evans, Dumortiera hirsuta (Sw.) Nees, Jamesoniella autumnalis (DeCandolle) Steph., Micromitrium austini Sull. in Aust., Notothylas breutelii (Gottschke) Gottschke, Riccardia multifida (L.) Gray ssp. synoica R.M. Schust.). Further investigation is required to verify the status of these identifications and current known distributions. Although not new to the state, the collections of Anacamptodon splachnoides (Froel. ex Brid.) Brid., Pogonatum pensilvanicum (Hedw.) Paris, and Syrrhopodon parasiticus (Sw. ex Brid.) represent one of the few (<5) vouchers of these species in the state.

None of the taxa on the current checklist are listed as endangered or threatened in Texas (TPWD 2011). NatureServe places several taxa in the “imperiled” or “vulnerable” categories at the global level (NatureServe 2011). However, a strict interpretation of these rankings is premature since assignment of these rankings is complicated by uncertainty in the currently accepted taxonomy, and insufficient distribution data. As a result, several of the taxa are assigned a status which varies widely from “vulnerable” to “secure.” The only taxon that would appear to be truly imperiled is Riccardia cf. jugata R.M. Schust., and this identification is at this point considered tentative.

**DISCUSSION**

Public interest in the flora of a region is often focused on the more obvious components of the vegetation - the vascular plants. These organisms, generically referred to as trees, wildflowers, weeds, etc., typically dominate the landscape in most ecosystems in Texas. In contrast, the lower plants, the group to which bryophytes are assigned, are often less conspicuous members of many ecosystems. Eula Whitehouse, in her brief synopsis of the state of bryology in Texas (Whitehouse 1952), references statements by early bryologists suggesting Texas was “almost unexplored” relative to bryophytes. Although the ensuing decades have increased our knowledge of these groups in the state, there is ample evidence to suggest there is more to be discovered.

The total number of species in this inventory represents a 57% increase in reported taxa since the initial BITH investigations completed by Mueller in 1974. The reasons for this increase are not clearly known. Additional collecting effort, defined as time spent in the field, coupled with a structured sampling scheme is one potential reason. Another possible explanation is an actual shift in species distributions resulting from changes in environmental conditions within a given region. The addition of the new species to the Texas bryoflora may be indicative of a shift in local conditions, albeit the directional nature of that shift is not clear. With only a few collections of specific taxa, the shift could be interpreted as immigration, or just as easily, as remnant species assemblages existing in highly specialized niches.

With little if any commercial value in Texas, bryophytes do nonetheless provide a number of important ecological benefits ranging from soil stabilization and nitrogen fixation, to bio-indicators of air and water.
pollution. As non-vascular plants, bryophytes are highly susceptible to changes in environmental conditions. This characteristic has led to a relatively recent utilization of bryophytes as early indicators of climate change (Tuba et al., 2011). In addition to movement of species to higher elevations, there is also an anticipated shift in species ranges toward the poles as the Earth’s climate warms. Due to the position of the BITH at the confluence of multiple eco-regions, and its location near the Gulf Coast of Texas, BITH could prove to be a natural laboratory for observing species, and community, distributional changes at both local and regional scales.

PRODUCTS

Two specimen record database files were developed and both have been forwarded to the NPS as part of this inventory. These two files contain records of specimens collected by the authors and also relevant records obtained in the herbaria search funded by the BTA. It is anticipated the results from this research will be published in the Journal of the Botanical Research Institute of Texas. The results of this inventory will also be included as part of a Ph.D. Dissertation by the first author.

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REFERENCES


Bazan, E. 1980. Hepatics of the Turkey Creek Unit of the Big Thicket National Preserve: A Floristic and Ecological Study. Master’s Thesis. Department of Biology, Texas A&M University, College Station, Texas.


Checklist of Bryophytes in the Big Thicket National Preserve, Texas

All taxa in this checklist were collected and identified by Dale A. Kruse or Paul V. Roling with the following exceptions. Taxa reported by previous authors and not collected in the current inventory are indicated as follows: (1) Bazan, 1980; (2) Mueller, 1974; (3) Stoneburner and Wyatt, 1979. Specimens located in herbaria and not collected in the current inventory are indicated using the appropriate herbarium acronym. Tentative state records are indicated with an asterisk. NatureServe rankings are included for taxa with a sole ranking of G3 or lower.

**ANTHOCEROTOPHYTA**

**Anthocerotaceae**  
*Anthoceros adscendens* Lehm. & Lindenb.\(^{(1)}\)  
*Anthoceros carolinianus* Michx.

**Notothyladaceae**  
*Notothylas breutelii* (Gottsche) Gottsche *\(^{*}\)

**MARCHANTIOPHYTA**

**Adelanthaceae**  
*Odontoschisma denudatum* (Nees Ex. Mart.) Dum. \(^{\text{(TAES)}}\)  
*Odontoschisma prostratum* (Sw.) Trev.

**Aneuraceae**  
*Aneura pinguis* (L.) Dumort  
*Riccardia cf. jugata* R.M. Schust. (G2)  
*Riccardia latifrons* (Lindb.) Lindb.\(^{(1)}\)  
*Riccardia multifida* (L.) Gray  
*Riccardia multifida* (L.) Gray ssp. synoica R.M. Schust. *\(^{*}\)

**Aytoniaceae**  
*Asterella elegans* ssp. *echinella* (Gott.) del Rosario \(^{\text{(TAES)}}\)  
*Asterella tenella* (L.) P. Beauv.\(^{(1)}\)  
*Reboulia hemisphaerica* (L.) Raddi

**Calypogeiaceae**  
*Calypogeia fissa* (L.) Raddi  
*Calypogeia sullivantii* Austin

**Cephaloziaceae**  
*Cephalozia catenulata* (Hüb.) Lindb.  
*Cephalozia connivens* (Dicks.) Lindb.  
*Cephalozia lunulifolia* (Dum.) Dum.

**Cephaloziellaceae**  
*Cephaloziella hyalina* Douin\(^{*}\)  
*Cylindrocolea obliqua* (Douin) Schust. (G1Q)  
*Cylindrocolea rhizantha* (Mont.) Schust.\(^{(1)}\)* (G3?)

**Fossombroniaceae**  
*Fossombronia brasiliensis* Steph.  
*Petalophyllum ralfsii* (Wilson) Nees & Gottsche ex Lehm.\(^{(2)}\)

**Geocalycaceae**  
*Lophocolea heterophylla* (Schrad.) Dum.

**Jubulaceae**  
*Frullania brtoniae* Evans \(^{(1)}\)  
*Frullania eboracensis* Gott.  
*Frullania inflata* Gott.  
*Frullania kunzei* Lehm. & Lindenb.  
*Frullania obcordata* Lehm. & Lindenb.  
*Frullania squarrosa* (Reinw., Blume & Nees) Dum.

**Jungermanniaceae**  
*Jamesoniella autumnalis* (DeCandolle) Steph. *\(^{*}\)  
*Jungermannia gracillima* Sm.\(^{(1)}\)*  
*Jungermannia hyalina* Lyell\(^{(1)}\)

**Lejeuneaceae**  
*Ceratolejeunea laetefusca* (Aust.) Schust. *\(^{*}\)  
*Cheilolejeunea rigidula* (Nees et Mont.) Schust.  
*Cololejeunea biddlecomiae* (Aust.) Evans \(^{(1)}\)*  
*Cololejeunea cardiocarpa* (Mont.) Schust.  
*Cololejeunea contractiloba* Evans *\(^{*}\)  
*Cololejeunea minutissima* (Sm.) Schiffn.  
*Cololejeunea setiloba* Evans *\(^{(2)}\)*  
*Lejeunea cladogyna* Evans (G3?)  
*Lejeunea flava* (Sw.) Nees  
*Lejeunea laetivirens* Nees et Mont.  
*Lejeunea ulcinia* (Tayl.) Gott.  
*Leucolejeunea clypeata* (Schwein.) Evans  
*Leucolejeunea conchifolia* Evans  
*Leucolejeunea unciolina* (Lindenb.) Evans  
*Leucolejeunea xanthocarpa* (Lehm. et Lindem.) Evans \(^{(2)}\)  
*Rectolejeunea maxonii* Evans

**Lepidoziaceae**  
*Kurzia sylvatica* (Evans) Grolle  
*Telaranea nematodes* (Gottsche ex Austin.) M. A. Howe
Marchantiaceae
Dumortiera hirsuta (Sw.) Nees

Metzgeriaceae
Metzgeria furcata (L.) Dumort.
Metzgeria uncigera A. Evans (1)* (G3)

Pallaviciniaceae
Pallavicinia lyellii (Hook.) Carruth.

Plagiochilaceae
Plagiochila dubia Lindenb. et Gott. (1)
Plagiochila ludoviciana Sull.
Plagiochila miradorensis Gott.

Porellaceae
Porella pinnata L.
Porella platyphylla (L.) Pfeiff.
Porella platyphyloidea (Schwein.) Lindb. (2)

Radulaceae
Radula australis Aust.
Radula obconica Sull.

Ricciaceae
Riccia curtisii (Aust.) James (1)
Riccia fluitans L. (2)
Riccia membranacea Gott. et Lindenb.
Riccia sorocarpa Bisch.
Riccia stenophylla Spruce
Riccia sullivantii Aust.
Ricciocarpos natans (L.) Corda

Scapaniaceae
Scapania nemorosa (L.) Dum.
Scapania nemorosa fo. whitehouseae R.M. Schust.

Sphaerocarpaceae
Sphaerocarpos texanus Aust.

BRYOPHYTA

Amblystegiaceae
Amblystegium serpens (Hedw.) Schimp. in B.S.G.
Amblystegium varium (Hedw.) Lindb.
Campylium chrysophyllum (Brid.) J. Lange
Leptodictyum riparium (Hedw.) Warnst.

Anomodontaceae
Anomodon attenuatus (Hedw.) Hüb.
Anomodon minor (Hedw.) Fürnr.

Bartramiaaceae
Philonotis longiseta (Michx.) E. Britton

Brachytheciaceae
Brachythecium acuminatum (Hedw.) Aust. (TAES)
Brachythecium bivertosum (Müll. Hal.) A. Jaeger (2)

Bryaceae
Bryum cf. radiculosum Brid.
Bryum pseudotriquetrum (Hedw.) Gaertn., Mey. & Scherb.

Calympereaceae
Syrrhopodon parasiticus (Brid.) Besch.
Syrrhopodon texanus Sull.

Climaceaceae
Climacium americanum Brid.

Cryphaeaceae
Cryphaea glomerata Bruch & Schimp. ex Sull.

Dicranaceae
Dicranella heteromalla (Hedw.) Schimp. (2)
Dicranella varia (Hedw.) Schimp. (2)
Dicranum condensatum Hedw. (2)

Ditrichaceae
Ceratodon purpureus (Hedw.) Brid. (TAES)
Ditrichum pallidum (Hedw.) Hampe

Entodontaceae
Entodon macropodus (Hedw.) C. Müll.
Entodon seductrix (Hedw.) C. Müll.

Ephemereaceae
Ephemera spinulosa Bruch & Schimp. in Schimp.
Micromitrium austini Sull. in Aust.*
Fabroniaceae
Anacamptodon splachnoides (Fröl. ex Brid.)
Brid.
Clasmatodon parvulus (Hampe) Hook. & Wilson
ex Sull. in Gray
Fabronia ciliaris (Brid.) Brid.

Fissidentaceae
Fissidens adiantoides Hedw.
Fissidens bryoides Hedw.
Fissidens bushii (Card. & Thér.) Card. & Thér.
Fissidens dubius P. Beauv.
Fissidens fontanus (B. Pyl.) Steud.
Fissidens polypodioides Hedw.
Fissidens ravenelii Sull.
Fissidens subbasilaris Hedw.
Fissidens taxifolius Hedw.

Fontinalaceae
Fontinalis novae-angliae Sull.

Funariaceae
Funaria americana Lindb. in Sull. (G3?)
Funaria flavicans Michx.
Funaria hygrometrica Hedw.
Funaria serrata Brid.
Physcomitrium immersum Sull.
Physcomitrium pyriforme (Hedw.) Hampe

Hypnaceae
Ctenidium molluscum (Hedw.) Mitt.
Herzogiella turfacea (Lindb.) Ivats. (G2)
Homomallium adnatum (Hedw.) Broth.
Hypnum lindbergii Mitt.
Isopterygium tenerum (Sw.) Mitt.
Platygyrium repens (Brid.) Schimp. in B.S.G. (G2)
Pseudotaxiphyllum homomallifolium (Redf.)
Irel. (G2G4)
Taxiphyllum taxirameum (Mitt.) Fleisch. (SHST)

Leptodontaceae
Forststroemia trichomita (Hedw.) Lindb.

Leskeaceae
Bryohaplocladium microphyllum (Hedw.)
Wat. & Ivats.
Leska australis Sharp
Leska gracilescens Hedw.
Leska obscura Hedw.
Leskea polycarpa Hedw.

Leucobryaceae
Leucobryum albidum (Brid. ex P. Beauv.) Lindb.
Leucobryum antillarum Schimp. ex Besch. (G2)
Leucobryum glaucum (Hedw.) Ångstr. in Fries.

Leucodontaceae
Leucodon julaceus (Hedw.) Sull.

Mniaceae
Mnium affine Bland. ex Funck.
Plagiochnium ciliare (C. Müll.) T. Kop.
Plagiochnium ellipticum (Brid.) T. Kop. (G2)
Rhizomnium punctatum (Hedw.) T. Kop.

Myriaceae
Schwetschkeopsis fabronia (Schwägr.) Broth.

Orthotrichaceae
Orthotrichum diaphanum Brid. (G2)
Orthotrichum obtusifolium Brid. (G2)
Schlotheimia rugifolia (Hook.) Schwägr.

Polytrichaceae
Atrichum angustatum (Brid.) Bruch & Schimp. in B.S.G.
Atrichum tenellum (Röhl.) Bruch & Schimp. in B.S.G.
Atrichum undulatum (Hedw.) P. Beauv.
Pogonatum brachyphyllum (Michx.) P. Beauv.
Pogonatum undulatum (Hedw.) P. Beauv.
Polytrichum commune Hedw.
Polytrichum ohioense Ren. & Card. (G2)

Pottiaceae
Astomum ludovicianum (Sull.) Sull.
Barbula indica (Hook.) Spreng. in Steud.
Barbula unguiculata Hedw.
Tortella fragilis (Hook. & Wils. in Drumm.)
Limpr. (SHST)
Tortella humilis (Hedw.) Jenn.
Tortula pagorum (Milde) De Not.
Weissia controversa Hedw.

Ptychomitriaceae
Ptycomitrium drummondi (Wils.) Sull.

Sematophyllaceae
Brotherella recurvans (Michx.) Fleisch. (G3)
Sematophyllum adnatum (Michx.) Britt.
Sphagnaceae
*Sphagnum affine* Ren. & Card.
*Sphagnum henryense* Warnst.
*Sphagnum lescurii* Sull. in Gray
*Sphagnum macrophyllum* Brid.
*Sphagnum molle* Sull.
*Sphagnum palustre* L.
*Sphagnum perichaetiale* Hampe

Theliaceae
*Thelia hirtella* (Hedw.) Sull. in Sull. & Lesq.
*Thelia lescurii* Sull. in Sull. & Lesq.

Thuidaceae
*Cyrto-hypnum minutulum* (Hedw.) Buck & Crum
*Thuidium allenii* Aust.
*Thuidium delicatulum* (Hedw.) Schimp. in B.S.G.
*Thuidium recognitum* (Hedw.) Lindb.(2)